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#### New feature!

es is also visible in a mutant deficient in mich extraplastidic lipids are less 1986a). Thus, the interaction between the assembly is complex—involving intricate al. 1986a). pid assemt bid assembly is complex—involving intricate indicate. I diadace, i disinguishing liquid assembly and transport become cc. expetually more difficult with the discovery of ER-plastid contact sites (Figure 3a,b) that give rise to a plastid-associated microsome fraction (PLAM) with classically ER-associated enzymatic Interview in a count (r-t-wny with classically EK-dssociated enzymatic activities (Kjeliberg et al. 2000). These contact sites have recently been visualized, and the strength of interaction between the two membrane systems has been determined using optical tweezers (Andersson et al.

÷ LIPID TRANSFER BETWEEN THE ER AND PLASTID ENVELOPES IPID TRANSFER BETWEEN THE ER AND PLASTID ENVELOPES

The complexity and redundancy of thylakoid lipid biosynthesis as depicted
in Figure 3b clearly invokes lipid transport processes that must shuttle
lipid precursors and products between the three involved biogenic
membranes and the thylakoids. In recent years, *Arabidopsis* genetics
provided identification of some of the genes and proteins involved in the
process, and a current model depicing the location and possible function
of these proteins is shown in Figure 4. In addition to the already
mentioned ats' mutants of Arabidopsis were identified that clearly meet
biosynthesis. These are the tgy mutants and arabidopsis were identified that clearly meet
oligogalactoglycerolipids, for example, tingalactosyldiactylycerolipids
curded the tipsical estructurally different from the typical 2007; Xu et al. 2003, 2006, 2008a). The oligogalactogycerolipids produced in the tdp mutants are structurally different from the typical galactoglycerolipids found in leaves, such that they are not likely the product of the nonprocessive UDP-Gai-dependent MGD1 or OED1 galactosyltransferases or their MGD2/3 and OGD2 paralogs (Xu et al. 2003). Instead, these oligogalactoglycerolipids appear to be produced by a processive UDP-Gai-independent galactosyltransferase associated with the outer envelope membrane. This activity was also perviously observed in plastid preparations by Wintermans and colleagues (Heemskerk et al.

FIGURES REFERENCES RELATED REVIEWS KEYV Andersson MX, Goksor M, Sandelius AS. 2007. Optical manipulation reveals strong attracting forces at membrane contact sites between endoplasmic reticulum and chioroplasts. J. Biol. Chem. 252(2):1170–144
 First detailed characterization of ER plastid outer envelope contact sites. Refl [Medline] [ISI]

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P. 2. Andersson MX, Kjellberg JM, Sandelius AS. 2001. Chloroplast biogenesis. Regulation of lipid transport to the thylakoid in chloroplasts isolated from expanding and fully expanded leaves of pea. Plant Physiol. 127(1):184–93 [CrossRef] [Medine] [ISI]

I 3. Andersson MX, Larsson KE, Tjelistrom H, Liljenberg C, Sandelius AS, 2005. Phosphate-limited oat. The plasma membrane and the tonoplast as major targets for phospholipid-0-glycolipid replacement and stimulation of phospholipases in the plasma membrane. J. Biol. Chem. 280(3):27578–86 [CrossRef] [MedInfe

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